Neuroprotective activity of Ethanolic Extract of *Tinospora cordifolia* on LPS induced Neuroinflammation


Department of Pharmacology, KK College of Pharmacy, Gerugambakkam, Chennai, India

*Corresponding author: Dr. Prakash R
prakasheeba@rediffmail.com

Department of Pharmacology, KK College of Pharmacy, Gerugambakkam, Chennai-600128, India.

Tel: 9940634459


Abbreviations

LPS: Lipopolysaccharide; EETC: Ethanolic Extract of *Tinospora cordifolia*; TLR: Toll-Like Receptor; NSAID: Non-Steroidal Anti-Inflammatory Drug; GSH: Glutathione Reductase; SOD: Superoxide Dismutase; TBARS: Thiobarbituric Acid Reactive Substances; CAT: Catalase; ROS: Reactive Oxygen Species

Introduction

Lipopolysaccharide (LPS), also known as bacterial endotoxin are biologically active substances, a major cell constituent present on the outer membrane of the cell wall of most Gram-negative bacteria [1,2]. Bacterial lipopolysaccharide consists of a polysaccharide region that is anchored on the outer bacterial cell wall.
membrane by a specific carbohydrate lipid moiety termed as Lipid A. This lipid A is responsible for the potent immunostimulatory property of LPS [3,4]. LPS projects its potency and act as an endotoxin by its greater affinity towards Toll-like receptor 4 (TLR 4) [5]. TLRs initiate key inflammatory responses and also shape adaptive immunity. LPS binds to TLR 4 which interacts with three different extracellular proteins namely LPS-binding protein (LBP), CD14 (Pattern recognition receptor-cluster of differentiation 14), Myeloid differentiation protein 2 (MD-2), to induce a signaling cascade, leading to activation of NF-κB-dependent induction of pro-inflammatory cytokines such as TNF-α, IL-1β, IL-6, COX, Prostaglandins and free radicals [3,6-8]. Thus, it is showed that by promoting the formation of free radicals, cytokines and other pro-inflammatory mediators in brain, blood as well as in tissue. LPS acts as a strong stimulator of potent immune system and produces a potent immunostimulant effect. As an outcome of potent stimulation of immune system by LPS, there results neuroinflammation.

Currently the term “Neuroinflammation” is used to describe the inflammatory response originated in the CNS due to the accumulation of glial cells by activating the immune component. But currently there are drugs available for the treatment of neuroinflammation which includes classes of drugs like NSAIDS, Opioid antagonists, selective COX inhibitor, NMDA receptor antagonists and rarely antibiotics [9,10]. But these allopathic medicines on usage, although effective at relieving pain and inflammation, produce undesirable serious side effects. For example, NSAIDS causes gastric irritation, abdominal or stomach pain, nausea, cramping, ringing in ears, confusion etc [11]. On the other hand, Opioid antagonists also results in unwanted effects like loss of appetite, dizziness, nervousness, etc. [12]. As an alternative method of treatment, in order to avoid all major side effects, we have focused our research work on Traditional medicines. These herbal medicines can be used to treat patients with lesser or no side effects as with allopathy.

_Tinospora cordifolia_ (wild) is used in ayurvedic “Rasayanas”, is a large, glabrous, deciduous, climbing shrub belonging to the family Menispermaceae indigenous to the tropical areas of India, Myanmar and Srilanka [13,14]. It is widely used in veterinary folk medicine, ayurvedic system of medicine for its general tonic, anti-diabetic, antipyretic, antispasmodic, anti-arthritic, antioxidant, anti-allergic, anti-stress, anti-inflammatory etc [15-18]. In the present study, we have investigated the effect of Extract of Tinospora cordifolia (EETC) which attenuates LPS induced neuroinflammation by down-regulating the levels of pro-inflammatory cytokines such as TNF-α, IL-1β, IL-6, COX, NOS etc.

Till date, no research work is scientifically validated on _Tinospora cordifolia_ on LPS induced neuroinflammation in rats. The present investigation was therefore aimed at evaluating the neuroprotective effect of _Tinospora cordifolia_ against LPS induced sickness behavior in rats.

### Materials and Methods

#### Authentication of plant material

The flower part of the plant _Tinospora cordifolia_ (Menispermaceae) was collected from Andhra Pradesh. The plant material was identified and authenticated by Dr. D. Narasimhan, Associate Professor, and Centre for Floristic Research, Madras Christian College, Tambaram, Chennai.

#### Preparation of extract: The fresh flower was collected and washed, chopped, dried at room temperature, and was made into coarse powder. The powder was extracted with 98% ethanol (68°C-78°C) in Soxhlet apparatus for 24 h. The extracts were concentrated on water bath (50°C). The yield of the ethanolic extract was found to be 1.0% (w/w) which was then stored at room temperature.

#### Animals

Adult male Wistar rat (160-180 g: 30 rats) were procured from CL Baid Metha College of Pharmacy, Chennai and divided into five groups of six animals each. The rats were housed in colony cages at an aberrant temperature of 25°C ± 2°C with a 12 h light/dark cycle. The animals had free access to standard pellet diet and drinking water. Behavioral studies were carried out in a quiet room between 9.00 am and 11.00 am to avoid circadian variation. The study was approved by Institutional Animal Ethical Committee, and work was carried out as per CPCSEA Guidelines, New Delhi.

#### Drugs and chemicals

Lipopolysaccharide (Sigma Aldrich, Mumbai), Aspirin (Madras Pharmaceuticals Ltd., Chennai), Thiobarbituric acid (S.d. Fine Chemicals Ltd., Mumbai), and SOD (HiMedia Research Laboratory, Mumbai) were obtained. All other chemicals and reagents unless specified were of analytical grade.

#### Experimental Design

The animals are divided into five groups with six rats each. The first group (control) received normal saline (0.9% NaCl, p.o.) once daily for 14 days. The second group treated saline for 14 days followed by LPS (100 μg/kg, i.p.). The third group received Aspirin (200 mg/kg p.o) for 14 days followed by LPS (100 μg/kg i.p.). The fourth group received EETC (200 mg/kg p.o) for 14 days followed by LPS (100 μg/kg i.p.). The fifth group received EETC (400 mg/kg p.o) for 14 days followed by LPS (100 μg/kg i.p.). All drugs were prepared freshly and given once daily in the morning and followed the same regimen (Table 1).

#### Table 1 Treatment according to group.

<table>
<thead>
<tr>
<th>Groups</th>
<th>Treatment</th>
</tr>
</thead>
<tbody>
<tr>
<td>Group-I</td>
<td>Normal saline (0.9% NaCl i.p.)</td>
</tr>
<tr>
<td>Group-II</td>
<td>Normal saline + LPS (100 μg/kg i.p.)</td>
</tr>
<tr>
<td>Group-III</td>
<td>Aspirin (200 mg/kg p.o) + LPS (100 μg/kg i.p.)</td>
</tr>
<tr>
<td>Group-IV</td>
<td>EETC (200 mg/kg p.o) + LPS (100 μg/kg i.p.)</td>
</tr>
<tr>
<td>Group-V</td>
<td>EETC (400 mg/kg p.o) + LPS (100 μg/kg i.p.)</td>
</tr>
</tbody>
</table>
Estimation of body weight

The body weights were measured before and after treatment with extracts and drug control. Body weight of all experimental animals was measured using a digital weighing scale.

Measurement of rectal temperature

Rectal temperature was measured with a thermostat probe inserted about 1.3 cm into the rectum and taped to the base of the tail. Temperature was monitored continuously.

Behavioral assessment

Locomotor activity: The spontaneous locomotor activity was monitored using digital Actophotometer equipped with infrared sensitive photo cells. The apparatus was placed in a darkened, light and sound attenuated, and ventilated testing room. Each interruption of beam on the x or y axis generated an electric impulse, which was presented on a digital counter. The activity was performed on day 14th on animals following LPS administration and the respective values were expressed as counts per 5 min [19].

Step through passive avoidance test

The step through passive avoidance apparatus consisted of an illuminated chamber (11.5 cm Â 9.5 cm Â 11 cm) attached to a darkened chamber (23.5 cm Â 9.5 cm Â 11 cm) containing a metal floor that could deliver foot shocks. The two compartments were separated by a guillotine door. The illuminated chamber was lit with a 25 W lamp. Briefly, rat was placed in the dimly lit room containing the apparatus 0.5 h before training to acclimatize to the new environment. Each rat was then placed individually into the illuminated chamber, facing away from the door to the dark chamber, and allowed to acclimatize for 1 min. As soon as the rat entered the dark chamber, the door was slid back into place, triggering a mild foot shock (0.3 Ma, 50 Hz, 5 s). The rat was then immediately removed from the chamber and returned to its home cage. The latency (time used to change compartment) was recorded. The retention test was conducted 24 h later (day 15) with the rat again being placed in the illuminated chamber and subjected to the same protocol in the absence of foot shock. The upper time limit was set at 300 secs [20].

Dissection and homogenization

After the treatment period, animals were sacrificed on 16th day by decapitation under mild anaesthesia. The brains were immediately removed, forebrain was dissected out, and cerebellum was discarded. Brains were put on ice and rinsed in ice-cold isotonic saline to remove blood. A 10% (w/v) tissue homogenate was prepared in 0.1 M phosphate buffer (pH 7.4). The homogenate was centrifuged at 10,000 g for 15 minutes and aliquots of supernatant obtained were used for biochemical estimation.

Estimation of glutathione reductase (GSH)

The reaction mixture containing 1 mL of phosphate buffer, 0.5 mL of ethylenediamine tetraacetic acid (EDTA), 0.5 mL of oxidized glutathione and 0.2 mL of NADPH was made up to 3 mL with distilled water. After the addition of 0.1 mL of tissue homogenate, the change in optical density at 340 nm was monitored for 2 min at 30 s intervals. One unit of the enzyme activity was expressed as moles of NADPH oxidized/min/mg protein [21].

Estimation of superoxide dismutase (SOD)

The supernatant (500 µL) was added to 0.8 ml of carbonate buffer (100 mM, pH 10.2) and 100 µL of epinephrine (3 mM). The change in absorbance of each sample was then recorded at 480 nm in spectrophotometer for 2 min at an interval of 15 s. Parallel blank and standard were run for determination of SOD activity. One unit of SOD is defined as the amount of enzyme required to produce 50% inhibition of epinephrine auto-oxidation. The reaction mixtures are diluted 1/10 just before taking the readings in the spectrophotometer [22].

Assay of thiobarbituric acid reactive substances (TBARS)-malonaldehyde level

The amount of malonaldehyde was used as an indirect measure of lipid peroxidation and was determined by reaction with thiobarbituric acid (TBA) [23]. Briefly, 1 mL of aliquots of supernatant was placed in test tubes and added to 3 mL of TBA reagent: TBA 0.38% (w/w), 0.25 M hydrochloric acid (HCl), and trichloroacetic acid (TCA 15%). The solution was shaken and placed for 15 min, followed by cooling in an ice bath. After cooling, solution was centrifuged to 3500 g for 10 min. The upper layer was collected and assessed with a spectrophotometer at 532 nm. Results were expressed as nanomoles per mg of protein. The concentration of MDA was calculated per mg of protein.

Estimation of catalase (CAT) level

The catalase activity was assessed by the method of Aebi [24]. The assay mixture consists of 0.05 mL of supernatant of tissue homogenate (10%) and 1.95 mL of 50 mM phosphate buffer (pH 7.0) in 3 mL cuvette. 1 mL of 30 mM hydrogen peroxide (H₂O₂) was added and changes in absorbance were followed for 30 s at 240 nm at 15 s intervals. The catalase activity was calculated using the millimolar extinction coefficient of H₂O₂ (90.071 mm mol cm⁻¹) and the activity was expressed as micromoles of H₂O₂ oxidized per minute per milligram protein.

Histopathological assessments

The brains from control and experimental groups were removed without any injury after opening the skull. The collected sample was washed with normal saline and fixed in 10% neutral formalin for 48 h for further histological observations. Hippocampus section was taken at 5 µm thickness processed in alcohol xylene series and was stained with crystal violet. The brain sections were examined microscopically for histopathological changes [25].

Statistical analysis

The results were expressed as mean ± SEM. Statistical differences between the mean of various groups were analysed by using variance followed by the Turkey multiple comparison test. Differences were considered significantly at P<0.05.
Results

Effect of EETC on LPS induced alterations in body weight

LPS (100 μg/kg) treated rats significantly (P<0.001) decreased in body weight when compared to control group. Aspirin (200 mg/kg) treated group showed significantly (P<0.001) increased in body weight compared to LPS treated rats. EETC (200 and 400 mg/kg) administered group significantly (P<0.001) increased the body weight when compared with LPS treated group (Figure 1).

Effect of EETC on LPS induced alterations in rectal temperature

LPS treated rats showed significantly (P<0.001) increased the rectal temperature when compared with control rats. Aspirin (200 mg/kg) treated group reversed the rectal temperature significantly (P<0.001) when compared with LPS treated group. In EETC (200 and 400 mg/kg) treated group reduced the rectal temperature significantly (P<0.001) when compared with LPS treated group (Figure 2).

Effect of EETC on LPS induced alterations in locomotor activity

LPS (100 µg/kg) treated rats significantly (p<0.001) decreased the locomotor activity in the actophotometer when compared to control rats. Pretreatment with Aspirin 200 mg/kg along with LPS significantly (p<0.001) increased the locomotor activity in the actophotometer when compared to LPS treated rats. Pretreatment with EETC (200 mg/kg) and EETC (400 mg/kg) significantly (p<0.001) increased the locomotor activity in the actophotometer when compared to LPS treated rats (Figure 3).

Effect of EETC on LPS induced memory impairment in passive avoidance test

The effect of EETC on long-term memory was investigated in the step-through passive avoidance test. During the training session (Day 1), there was no significant difference between any groups. However, there was a significant difference in the retention latency between the groups. LPS treated rats showed a significantly (P<0.001) lower latency when compared to control group which was performed 24 hrs after the training period. The reduced retention latency indicates learning and memory impairment in rats. This effect was reversed significantly (P<0.001) by administration of EETC 200 and 400 mg/kg. Aspirin treated group significantly increased the latency period during retention test when compared to LPS treated group. There was no significant difference in the retention latency period between the doses of EETC 200 mg/kg and EETC 400 mg/kg (Table 2).

Effect of EETC on anti-oxidant level

Effect of EETC on LPS induced alterations in GSH level: LPS treated rats significantly (P<0.001) decreased the level of GSH

![Figure 1](image1.png) Effect of EETC on LPS induced alterations in Body weight. Values are expressed mean± SEM. *P<0.001 compared to control rats.

![Figure 2](image2.png) Effect of EETC on LPS induced alterations in Rectal temperature. Values are expressed mean ± SEM. c P<0.001 and a P<0.05 compared to control rats. z P<0.001 compared to LPS treated rats.

![Figure 3](image3.png) Effect of EETC on LPS induced alterations in Locomotion using actophotometer. Values are expressed mean± SEM. c P<0.001 and a P<0.05 compared to control rats. z P<0.001 compared to LPS treated rats.
when compared to control rats. Aspirin treated rats significantly (P<0.001) increased the GSH level when compared with the LPS treated rats. EETC (200 and 400 mg/kg) treated rats increased the level of GSH produce significantly (P<0.001) when compared with LPS treated rats. There was no significant difference in the level of GSH between EETC 200 mg/kg and 400 mg/kg (Table 3).

Effect of EETC on LPS induced alterations in SOD level: LPS treated rats produced significantly (P<0.001) decrease in the level of SOD when compared with control rats. Aspirin treated group showed significantly (P<0.001) increased the level of SOD when compared with LPS treated group. EETC (200 and 400 mg/kg) restored significantly (P<0.001) the level of SOD but the better antioxidant effect was obtained from EETC 400 mg/kg than the EETC (200 mg/kg) (Table 3).

Effect of EETC on LPS induced alterations in TBARS level: LPS treated group significantly (P<0.001) increased the level of TBARS when compared with the control group. Aspirin treated group decreased the TBARS level significantly (P<0.001) when compared with LPS treated rats. EETC (200 and 400 mg/kg) administered group significantly (P<0.001) decreased the level of TBARS when compared to LPS treated group but the effect is less significant than Aspirin treated group. EETC 400 mg/kg administered group exerted its better anti-oxidant activity by decreased the level of TBARS when compared to EETC (200 mg/kg) (Table 3).

Effect of EETC on LPS induced alterations in CAT level: LPS treated group significantly (P<0.001) decreased the level of CAT when compared with control group. Aspirin treated group significantly (P<0.001) increased the CAT level compared to LPS treated rats. EETC (200 and 400 mg/kg) treated group significantly (P<0.001) increased the CAT level when compared to LPS treated group but less increase when compared to control and Aspirin treated group. There was no significant difference on the activity between the EETC (200 mg/kg) and EETC (400 mg/kg) treated groups (Table 3).

Histopathological assessment of EETC in hippocampus region using crystal violet staining: There was a decrease in the neuronal cells, increased in chromatolysis and pyknotic nuclei in LPS treated rats in the region of Hippocampal brain slices when compared to control rats. Aspirin treated rats showed an increase in the regeneration of neuronal cells compared to LPS rats. EETC (200 mg/kg) and EETC (400 mg/kg) treated rats showed significantly decrease in cell edema, neuronal cell degeneration compared to LPS treated rats by dose dependently (Figures 4a-4e).

### Discussion

The central findings of the current study reveal that EETC attenuates LPS induced sickness behavior on rats. LPS can bind with TLR4 leading to the activation of NF-κB-dependent induction of pro-inflammatory cytokines such as TNF-α, IL-1β, IL-6 and COX which leads to sickness behavior. These findings replicate the results of many previous studies, which demonstrated that activation of the immune system by LPS induces a Sickness behavior which refers to a coordinated set of behavioral changes that develop during the course of raised systemic inflammation [26] and include reduction in appetite and body weight, suppression of locomotor, exploratory, and social activity, fatigue and malaise, impairment in cognitive abilities, reduced libido and sexual behavior, anhedonia and impairment in motor behavior [27-29].

In our present study LPS treated rats showed decrease in body weight when compared to control rats due to stress. The body weight was increased to rats were treated with Aspirin and EETC (200 and 400 mg/kg) results from anti-stress activity with active food intake when compared to LPS treated group. The presence of alkaloids and diterpenoid lactone contents exert an anti-stress activity of EETC [30]. LPS produced a marked elevation in rectal temperature 1-3 h after i.p. administration. Some of the changes may found in neurotransmitter activities such as 5-HT and NE could account for this effect of LPS [31,32]. LPS produced a biphasic effect on body temperature, with an initial hypothermia, followed by a febrile phase where hyperthermia produced. The extract of T. cordifolia produced very good antipyretic effect in a dose-dependent manner and the observed

### Table 2

<table>
<thead>
<tr>
<th>Treatment</th>
<th>Acquisition Trail (Day 1)</th>
<th>Retention Latency (Day2)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>15.50 ± 1.08</td>
<td>78.67 ± 3.87</td>
</tr>
<tr>
<td>LPS (100 µg/kg, ip)</td>
<td>26.67 ± 0.66</td>
<td>24.17 ± 1.16 c</td>
</tr>
<tr>
<td>Aspirin (200 mg/kg, po)</td>
<td>16.33 ± 0.61</td>
<td>111.7 ± 4.82 z</td>
</tr>
<tr>
<td>EETC (200 mg/kg, po)</td>
<td>19.67 ± 1.20</td>
<td>97.33 ± 2.24 z</td>
</tr>
<tr>
<td>EETC (400 mg/kg, po)</td>
<td>18.33 ± 1.49</td>
<td>92.00 ± 1.69 z</td>
</tr>
</tbody>
</table>

Values are expressed mean ± SEM. *P<0.001 compared to control rats. **P<0.001 compared to LPS treated rats.

### Table 3

<table>
<thead>
<tr>
<th>Treatment</th>
<th>GSH (nmoles/g tissue)</th>
<th>SOD (units/min/mg protein)</th>
<th>TBARS (nmoles/mg protein)</th>
<th>CAT (mg protein/min)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>35.42 ± 6.42</td>
<td>8.12 ± 2.08</td>
<td>1.85 ± 0.12</td>
<td>3.98 ± 1.26</td>
</tr>
<tr>
<td>LPS (100 µg/kg, ip)</td>
<td>21.26 ± 2.49 c</td>
<td>3.92 ± 1.94 c</td>
<td>4.59 ± 0.96 c</td>
<td>1.43 ± 0.93 c</td>
</tr>
<tr>
<td>Aspirin (200 mg/kg, po)</td>
<td>32.18 ± 8.16 z</td>
<td>7.43 ± 2.96 z</td>
<td>1.98 ± 0.23 z</td>
<td>3.21 ± 1.34 z</td>
</tr>
<tr>
<td>EETC (200 mg/kg, po)</td>
<td>27.34 ± 6.32 x</td>
<td>5.97 ± 0.98 z</td>
<td>2.82 ± 0.46 y</td>
<td>2.29 ± 2.56 y</td>
</tr>
<tr>
<td>EETC (400 mg/kg, po)</td>
<td>29.31 ± 8.34 y</td>
<td>6.58 ± 1.02 z</td>
<td>2.12 ± 0.12 z</td>
<td>2.98 ± 1.45 z</td>
</tr>
</tbody>
</table>

Values are expressed mean ± SEM. *P<0.001 compared to control rats **P<0.001 and ***P<0.05 compared to LPS treated rats.
The antipyretic properties of *T. cordifolia* exerted due to the presence of berberine [33], diterpenoid lactones, aliphatic compounds and steroids [34]. The previous studies were showed the presence of water-soluble fractions of 95% ethanolic extract of *T. cordifolia* plant to possess significant antipyretic activity [35].

Following LPS injection in rats, a change in the total locomotion as depicted from decrease activity in Actophotometer was observed [36]. Administration of ethanolic extract of *T. cordifolia* and Aspirin increases the locomotion activity in rats compared to LPS treated group. The increase in the locomotion due to the presence of glycosidal contents in the plant [37] and thereby, suppresses the depression and anxiety in rats.

LPS treated rats produced cognitive and memory impairment shows decreased in latency period in passive avoidance test. The activation of inflammatory mediators upon microglial activation affects the hippocampus function [38]. Our present investigation exhibits anti-inflammation and cognitive stabilization by increasing retention latency period upon administration of EETC (200 and 400 mg/kg) and Aspirin (200 mg/kg). The EETC produces cognitive enhancement by the presence of glycosidal contents [37] and synthesis of acetylcholine. The increased level of choline shows memory enhancing property for learning and memory in normal and memory-deficits animals [39].

The tripeptide GSH is an important endogenous anti-oxidant which has a major role in restoring other free radical scavengers and anti-oxidants, such as vitamins C and E, to their reduced state [40]. The decreased level of GSH observed upon administration of LPS due to oxidative damage when compared to control group. The EETC (200 mg/kg and 400 mg/kg) treated rats restored the GSH level compared to LPS treated rats due to presence of highest phenolic contents [41] and extracts showing highest free radical scavenging activity. However, Aspirin significantly elevated the activities of antioxidative enzymes compared to LPS-treated animal.

Living organisms contain SOD, which removes superoxide, and are thus protected from injury caused by ROS [42]. A significant decrease in the enzyme activity of SOD was observed as a result of increased superoxide formation upon LPS administration at day 14 [36] due to oxidative stress. EETC produced positive modulatory effect on cellular anti-oxidant system by increasing the anti-oxidant level of SOD. The antioxidative activity of EETC (400 mg/kg) was potent than that of EETC (200 mg/kg). The increased level of SOD was observed on Aspirin treated rats when compared to LPS treated rats.

TBARS level elevated in the LPS administered rat due to oxidative stress of cells. The anti-oxidant property of Aspirin was enhanced by decreasing the formation of TBARS. The formation of TBARS in the brain was inhibited in a concentration dependent manner in presence of EETC, as the EETC (400 mg/kg) shows potent suppression of TBARS than EETC (200 mg/kg) in rats. This study supported from previous reports that, the (1,4)-alpha-d-glucan (alpha-d-glucan), derived from *T. cordifolia* have been shown to activate human lymphocytes with downstream synthesis of the pro- and anti-inflammatory cytokines, in vitro. Its ability to destroy cellular metabolic waste products may be responsible for the anti-oxidant effect and strengthens the tissues [43].

Catalase mediates its function by the removal of $H_2O_2$ generated by auto-oxidation of lipids and the oxidation of organic substances [36]. The study revealed that LPS treatment suppressed the CAT activities when compared to control group. The administration of EETC following LPS in rats reported to show scavenging activity due to the presence of antioxidants and had protective effect by increasing the CAT level. The anti-oxidant activity of Aspirin (200 mg/kg) treated rats showed by increase in the CAT level. The anti-oxidant property was higher in the dose of 400 mg/kg of EETC than in 200 mg/kg of EETC administered rats. The result supported from previous studies that, *T. cordifolia* contains Flavonoids, glycosides, saponins and some amount of phytosterols. These active constituents alone or in combination may be responsible for the observed antioxidant activity [44].

The LPS administration on rats caused neuronal injury induced cell damage on Hippocampal slices was observed clearly when compared with control because of microglial activation results in immunostimulation [45,46]. After treatment with the EETC showed increase in the regeneration of neuronal cells with composite nature and same process obtained from Aspirin treated group. This clearly showed that EETC protects the cells from get damage by built up of neuronal cell structure and reduces the cell edema. EETC 400 mg/kg dose expressed better protection when compared to 200 mg/kg of EETC and the results
obtained concludes the neuroprotective effect of EETC on brain from LPS inflammation.

Conclusion
The ethanolic extract of Tinospora cordifolia may elicit neuroprotective activity due to the presence of phytochemical constituents such as alkaloids, glycosides, diterpenoid lactones, berberine, flavonoids, saponins.

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References


