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In vitro antiviral activity of honey against varicella zoster virus (VZV): A translational medicine study for potential remedy for shingles



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Abstract

Objectives: The aim of this study was to determine the *in vitro* anti-viral effect of honey on varicella zoster virus.

Methods: Manuka and clover honeys were used at concentrations ranging from 0-6% wt/vol. A clinical VZV isolate was obtained from a zoster vesicle and used at low passage. Various concentrations of manuka and clover honey were added to the tissue culture medium of VZV-infected human malignant melanoma (MeWo) cells.

Results: Both types of honey showed antiviral activity against varicella zoster virus with an approximate EC50 = 4.5 % (wt/vol).

Conclusions: Our results showed that honey has significant *in vitro* anti-VZV activity. As, honey is convenient for skin application, is readily available and inexpensive, honey may be an excellent remedy to treat zoster rash in developing countries where antiviral drugs are expensive or not easily available.

Keywords: Varicella Zoster, Shingles, Zoster, Honey, Antivirals, Translational Medicine.

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Introduction

Herpes zoster (shingles) is the reactivation of varicella zoster virus from latency in cranial, autonomic and dorsal root ganglia and characterized by rash and severe pain in affected dermatomes. [1] Zoster is a disease affecting more than one million people per year in United States. [2] Zoster is especially common in developing poor countries possibly due to malnutrition, chronic diseases, and ineffective immunization programs. Patients affected by zoster in poor countries are more prone to develop complications such as postherpetic neuralgia (PHN), herpes zoster ophthalmicus, vasculitis, meningitis, and myelitis. [3] Antiviral VZV therapeutic agents effective for treating VZV reactivation such as acyclovir, famcyclovir and valacyclovir are very expensive in developing countries and are not readily available. There is great need for a remedy which is inexpensive and easily available in developing countries.

Since antiquity, honey has been used to treat many diseases. [4-8] Currently, honey has been shown to have excellent antibacterial activity for many wound pathogens. [9, 10] Honey has excellent antibacterial activity against Methicillin-resistant Staphylococcus aureus (MRSA) and various species of pseudomonas commonly associated with wound and burn infections. [11-14]Honey dressings are used commonly to manage skin and burn wounds infections. [15]Honeys also possess antifungal activity. [16] Honey has antiviral activity against Rubella virus [17], and honey is used topically to treat recurrent herpes simplex lesions. [18]In this study we screened two types of honey for antiviral activity against a clinical isolate of varicella zoster virus.

Materials & Method

Cell culture

Human malignant melanoma cells (MeWo) were grown at 37° C and 5% CO₂ in Dulbecco's minimal essential medium supplemented with 10% fetal bovine serum (DMEM, Life Technologies, Carlsbad CA). The MeWo cell line was established from a skin biopsy of malignant melanoma, and is the standard cell line used to propagate VZV.[19]

Virus stock

Varicella zoster virus (VZV) obtained from a zoster vesicle and shown to be of the European strain was used before the 20th *in vitro* passage [20] Since VZV is highly cell-associated, virus was propagated by co-cultivation of infected cells with uninfected cells at ratios ranging from 1:4 to 1:100 depending on use. VZV stocks were prepared using infection ratios of 1:100 and dose response experiments were performed at infection ratios of 1:4. [21].

Treatment with honey

Commercially obtained pure clover and Manuka honey were diluted to a final concentration ranging from 0% to 6% (wt/ vol) in culture medium and filter sterilized.

Cell viability assay

Figure 1. Viability of MeWo Cells in dilute

honey solutions.

Neutral red uptake assay was used to determine cell viability in various honey concentrations. MeWo cells

were incubated for 3 hrs with 40 ug/ ml neutral red in DMEM, fixed in 0.5% formaldehyde, 1% CaCl2 for 1 min,

at 560 nm on triplicate samples. Cytotoxicity for both honeys is similar until highest concentration tested. At 5

dissolved in 50% ethanol, 1% acetic acid for 5 min. Optical density was determined

% dilution, clover honey is more toxic to MeWo cells than Manuka honey.

Cell viability in various honey concentrations was determined by neutral red uptake assay. [22] Briefly, after 3 days in culture MeWo cells were incubated for 3 hrs with 40 ug/ml neutral red (Sigma, St. Louis, MO) in DMEM, fixed in 0.5% formaldehyde, 1% CaCl2 for 1 min, dissolved in 50% ethanol, 1% acetic acid for 5 min, and optical density at 560 nm determined on triplicate samples.

Infectivity assay

MeWo cells infected with VZV and uninfected MeWo cells were incubated in culture medium containing various honey concentrations. After plague development (3 days post-infection) cultures were fixed in 4% formaldehyde, permeabilized for 10 minutes in methanol/acetone (50:50; v/v) and extensively washed with Tris-buffered saline (TBS; 20 mM Tris-HCl, pH 8.0, 150 mM NaCl). VZV plaques were identified by immunostaining. Briefly, monolayers were blocked for 60 minutes in 3% blocked for 60 minutes in 3% BSA in TBS, incubated for 60 minutes with primary antibody (rabbit anti-IE63: 1:1,000 dilution) [23] followed by 60 minutes incubation with alkaline phosphatase conjugated secondary antibody (goat anti-rabbit IgG; 1:10,000 dilution; Abcam, Cambridge, MA). Immunostaining was visualized with NBT/BCIP (Pierce, Rockford IL). Between all incubations, the cultures underwent extensive washing in TBS. Plagues were counted with the aid of a dissecting microscope (4 x magnifications).

Results & Discussion

Both clover and Manuka honey are readily soluble in water, and at low concentrations inhibit neutral red uptake to equal amounts. MeWo cells in both honey concentrations \leq 3.75% showed equal survival slopes (**Fig. 1**). This is most likely due to



% survival of MeWo as a function of honey concentration

2

3

4

5

6

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- clover - Manuke Figure 2. Anti-VZV activity of manuka and clover honeys.

MeWo cells were infected with VZV and treated with the indicate concentrations of manuka and clover honeys. At 3 days post infection, virus plaques were immunostained and counted.







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120

100

80

60

40

20

0

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1

% survival

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nonspecific osmotic actions of the sugars. Both types of honey have concentration dependent *in vitro* anti-VZV activity with half maximal effective concentration (EC_{50}) approximately 4.5% (wt/vol) (**Fig. 2**). Manuka honey showed a slightly less EC_{50} as compared to clover honey. At the EC_{50} concentration, cells remained viable. Higher honey concentrations resulted in significant reduction in VZV plaque size. (**Fig. 3**)

For centuries, honey has been used in traditional medicine. In recent past, honey has gain significant attention from the scientific community to explore its potential applications to treat various clinical conditions. Honey has wide range of therapeutic properties including anti-inflammatory, antibacterila, antifungal and antineoplastic activity. [24, 25] Our results suggest the presence in honey of compounds possessing antiVZV activity, the identity of which is yet to be determined. Honey is convenient for application on skin, readily available and inexpensive, and potentially an excellent remedy to treat zoster rash in developing countries where antiviral drugs are expensive or not easily available.

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